



# 107° CONGRESSO NAZIONALE della SOCIETÀ ITALIANA DI FISICA



## Comparison of protein conformational properties *in solution and in the crystalline state* by Fourier transform infrared spectroscopy

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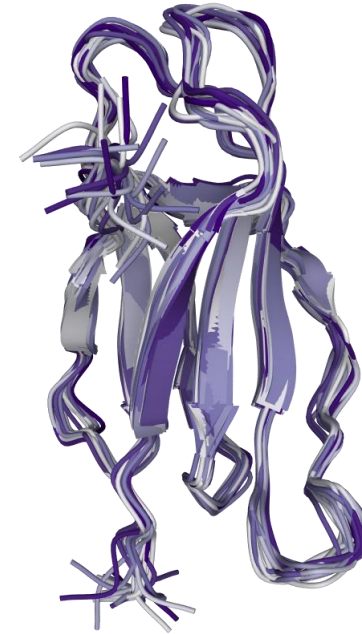
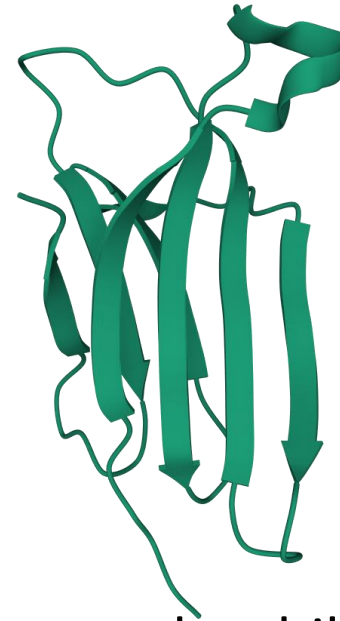
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# Proteins in solution and in the crystalline state

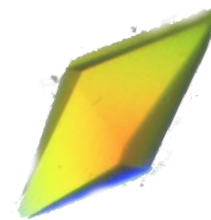
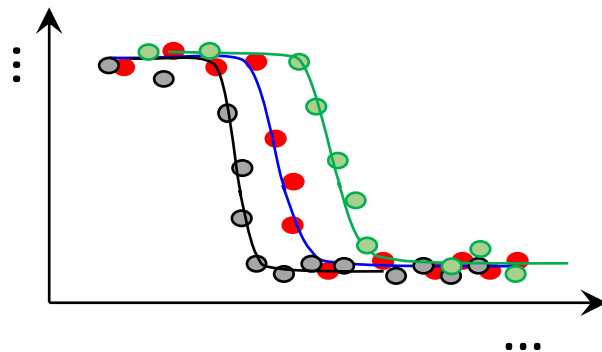
- ❖ Detailed 3D structures can be obtained both in solution and in the crystalline state

**1LDS**  
Crystal Structure of  
monomeric human  
beta-2-microglobulin  
DOI: [10.2210/pdb1LDS/pdb](https://doi.org/10.2210/pdb1LDS/pdb)



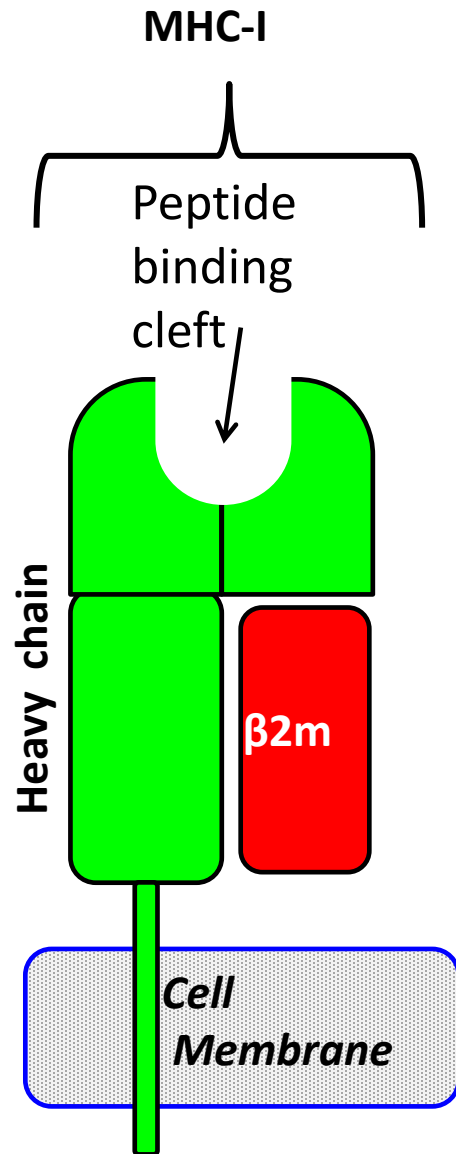
**1JNJ**  
NMR solution  
structure of the  
human  
beta2-microglobulin  
DOI: [10.2210/pdb1JNJ/pdb](https://doi.org/10.2210/pdb1JNJ/pdb)

- ❖ Protein conformational properties (i.e. structural stability and aggregation propensity) are typically studied in solution



- ❖ Are these “in solution” properties detectable in the protein crystals?
- ❖ How we can study these properties in the protein crystals?

# The model system: $\beta$ -2microglobulin



→  $\beta$ -2microglobulin ( $\beta$ 2m) is the light chain of class I major histocompatibility complex (MHC-I).

→ In vivo,  $\beta$ 2m is degraded and excreted by kidneys.

→ As a consequence of renal failure, the plasma concentration of  $\beta$ 2m increases up to 60-fold. This leads to the accumulation of  $\beta$ 2m around bones and joints in form of amyloid deposits, resulting in the development of the dialysis-related amyloidosis (DRA).

→ The natural occurring variant of  $\beta$ 2m D76N causes an autosomal dominant, hereditary systemic amyloidosis with extensive visceral amyloid deposits\*.

\*Valleix et al. 2012 N Engl J Med 366:2276-2283

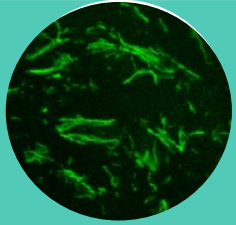
# $\beta$ 2-microglobulin is the causing agent of two types of systemic amyloidosis:

## Dialysis Related Amyloidosis

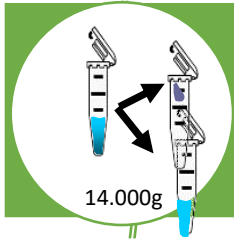
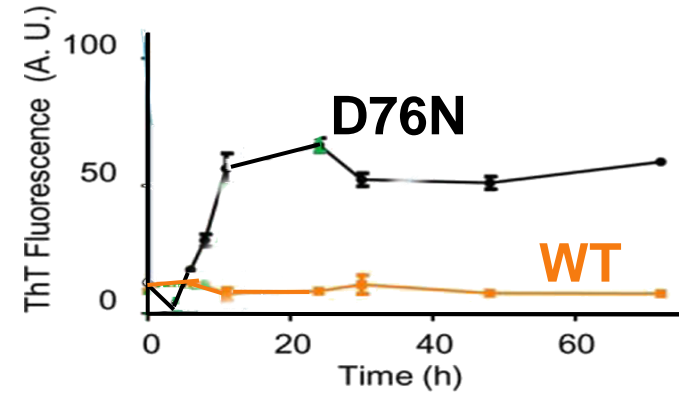
## D76N $\beta$ <sub>2</sub>m Amyloidosis

Acquired	←Type of amyloidosis→	Hereditary
After 10 years of Dialysis	←Onset→	After 50-60 years of life
50-70 mg/L	← $\beta$ <sub>2</sub> m plasma concentration→	2 mg/L
Osteotendinous tissue	←Site of deposition→	Systemic
Full length WT $\beta$ <sub>2</sub> m + $\Delta$ N6 $\beta$ <sub>2</sub> m 70%      30%	←Fibril composition→	Full-length D76N $\beta$ <sub>2</sub> m

Time course of aggregation studied by  
biophysical and biochemical approaches:  
PBS pH 7.4, 37°C, stirring.



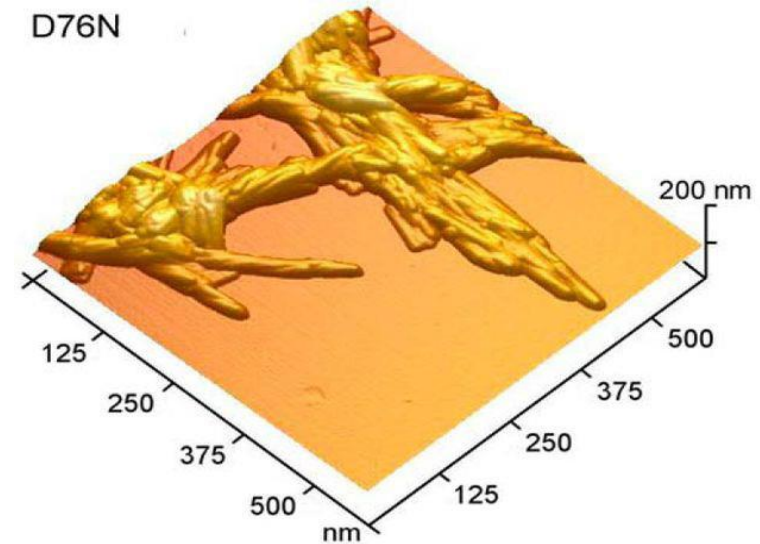
ThT assay: ThT binding is a common property of amyloid fibrils obtained from different precursor proteins



SDS-Page



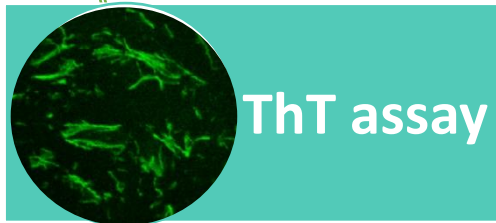
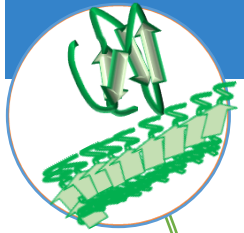
Microscopy: aggregate morphology at the nm scale



# Time course of aggregation studied by biophysical and biochemical approaches: PBS pH 7.4, 37°C, stirring.

## FTIR spectroscopy

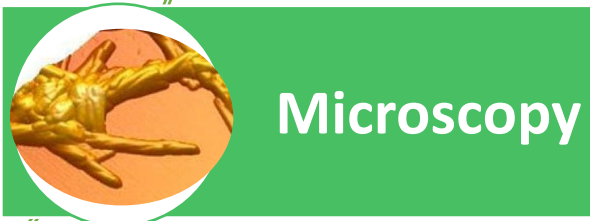
- ❖ It allows to study complex samples in different physical states (from soluble/insoluble proteins to intact cells and tissues)
- ❖ It allows discriminating native intramolecular  $\beta$ -sheets from intermolecular  $\beta$ -sheets



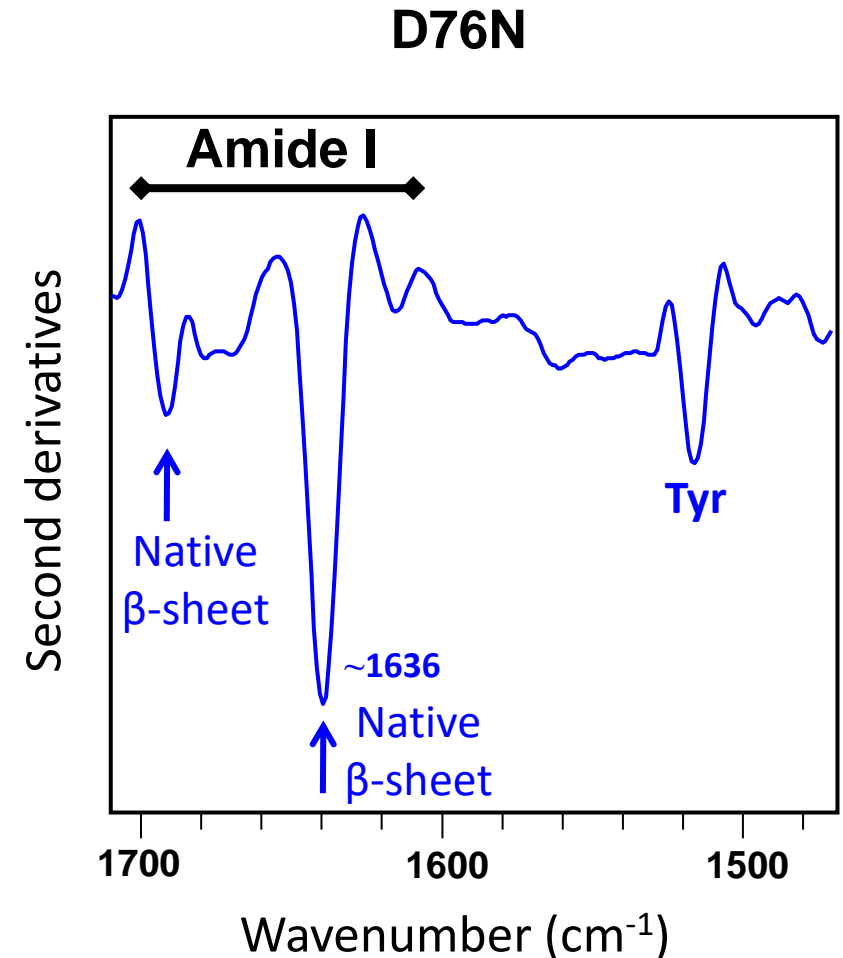
ThT assay



SDS-Page



Microscopy

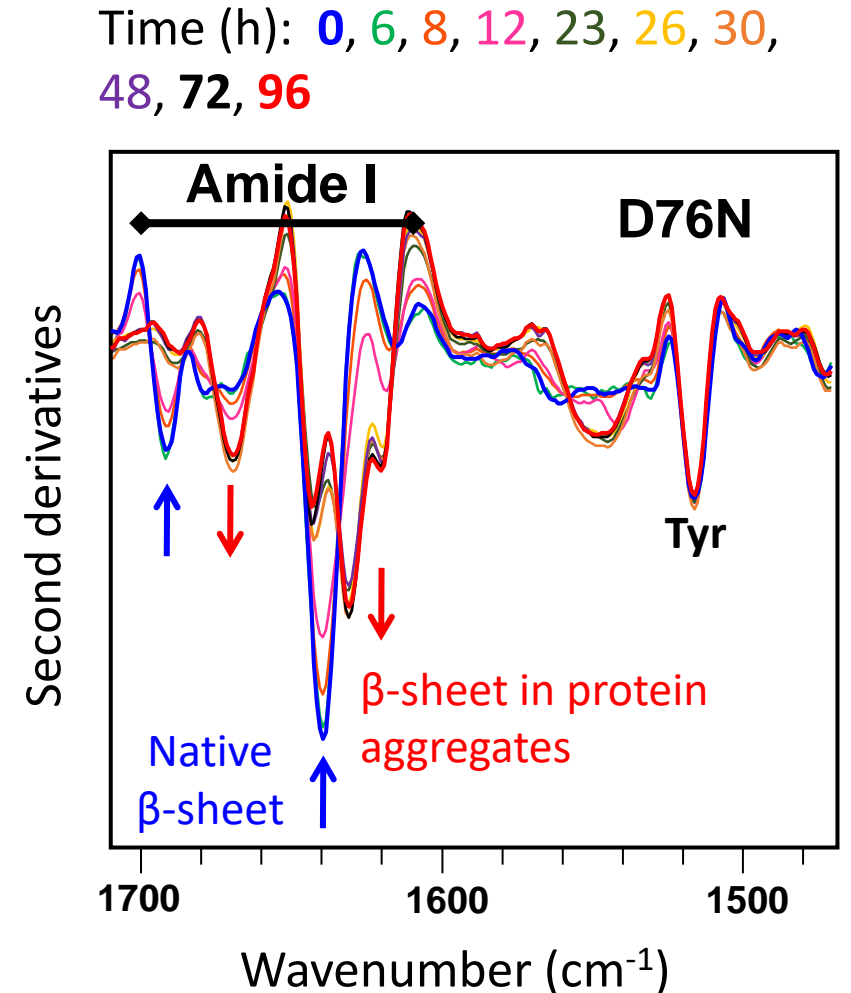
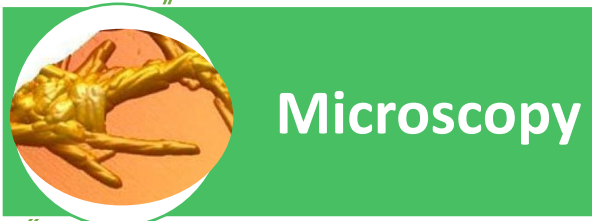
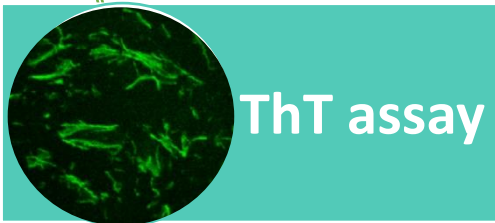
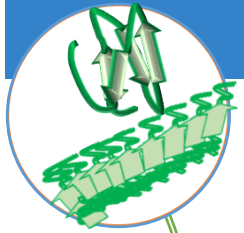




# Time course of aggregation studied by biophysical and biochemical approaches: PBS pH 7.4, 37°C, stirring.

## FTIR spectroscopy

- ❖ It allows to study complex samples in different physical states (from soluble/insoluble proteins to intact cells and tissues)
- ❖ It allows discriminating native intramolecular  $\beta$ -sheets from intermolecular  $\beta$ -sheets



## The model system: $\beta$ -2microglobulin variants

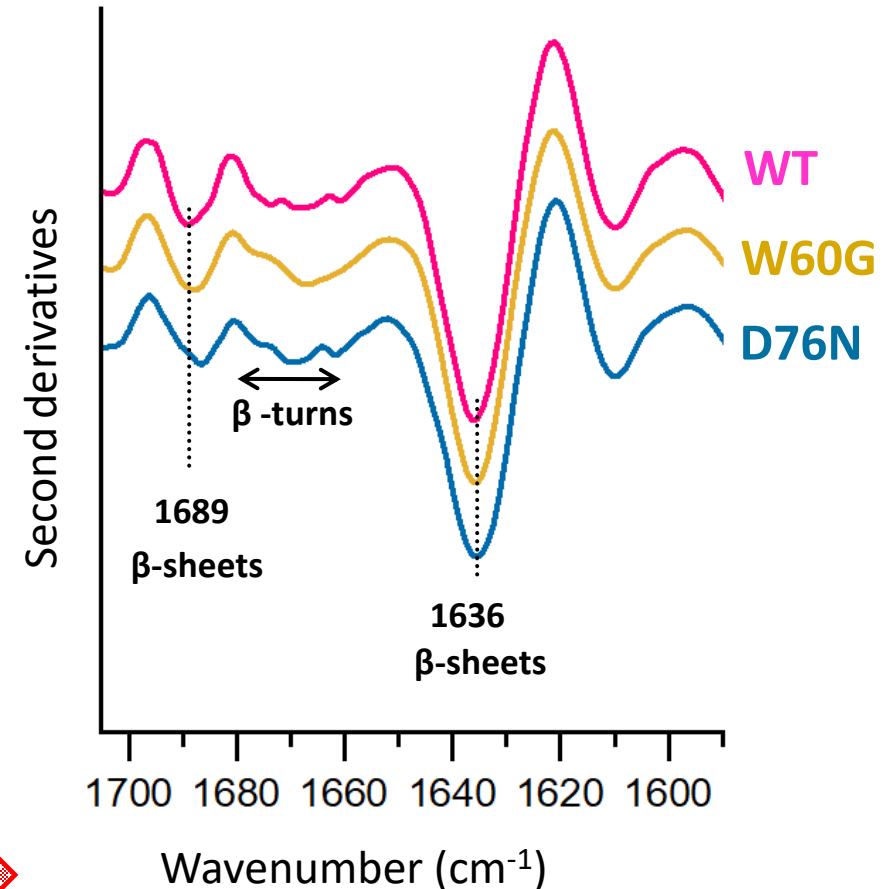
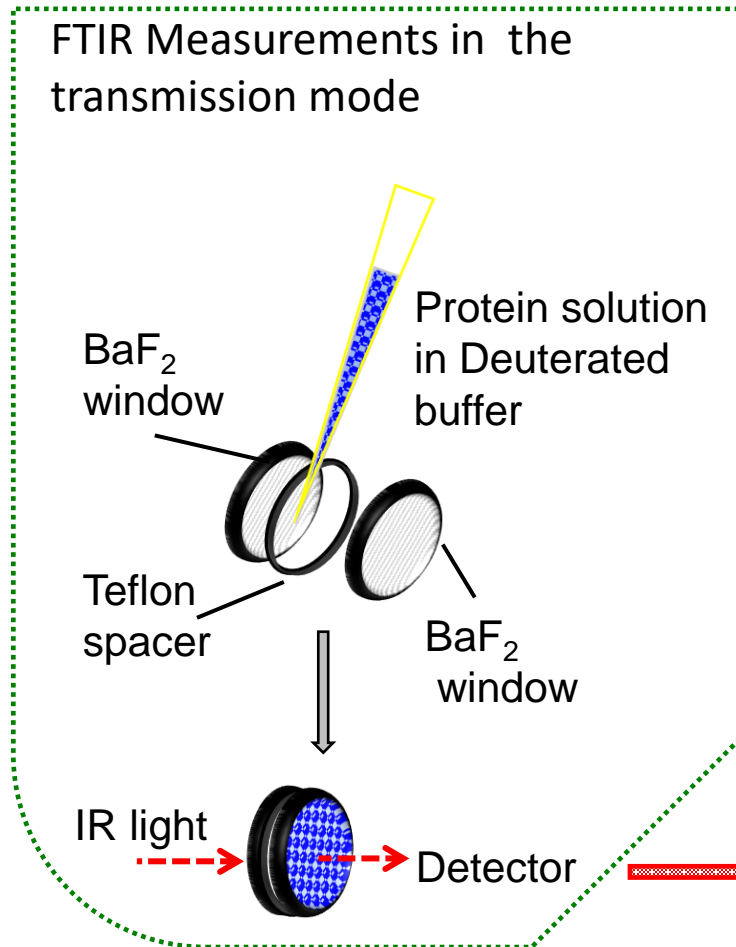


- ❖ In addition to the Amyloid-prone mutant (**D76N**), a stable variant was also considered for comparison: the **W60G** protein
- ❖ The structures of the **W60G** and **D76N** mutants are very similar to the structure of the **WT** protein.
- ❖ The three variants crystallize under identical conditions and form the intermolecular interactions in identical crystal packing.

Superposition of the crystal structure of human **WT** (PDB: **2YXF**)  $\beta$ 2m, **W60G** (PDB: **2Z9T**) and (**D76N** **4FXL**) mutants

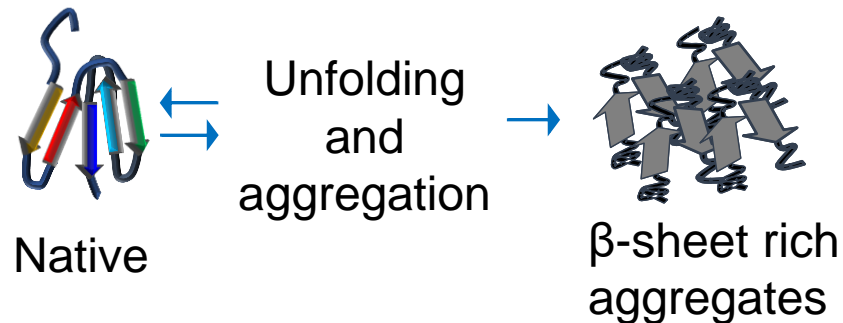
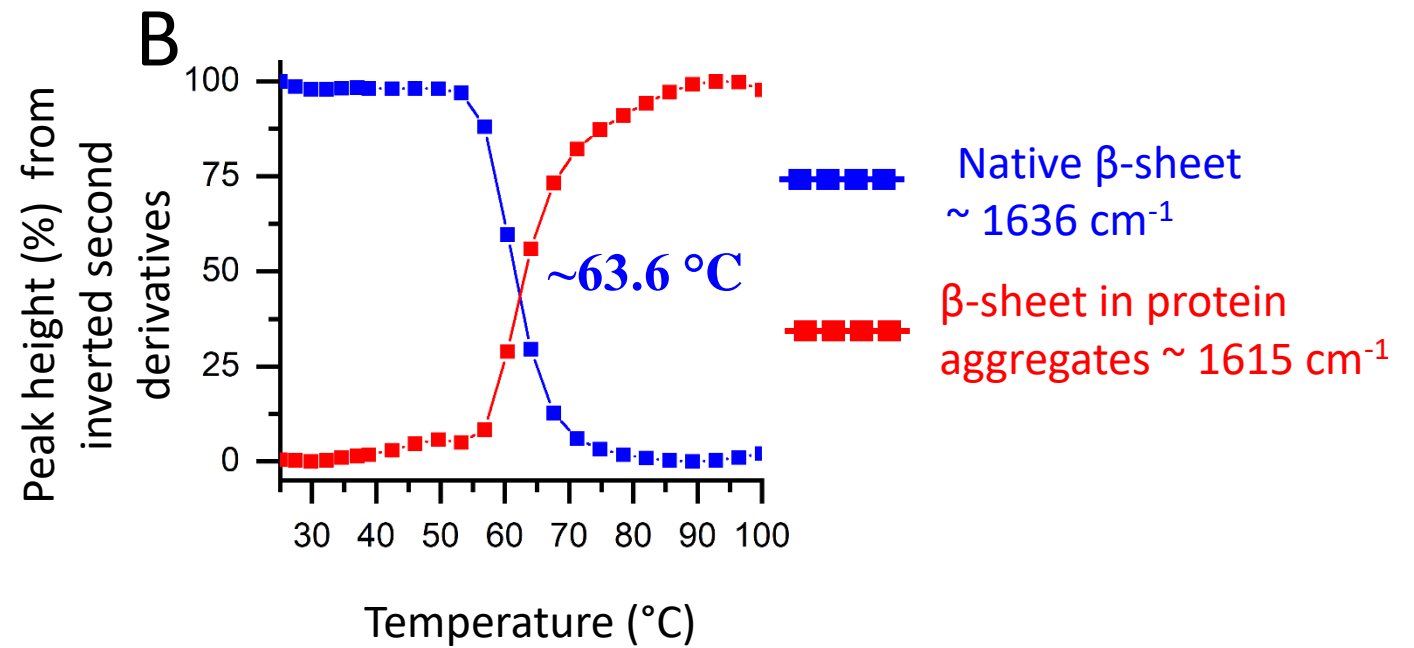
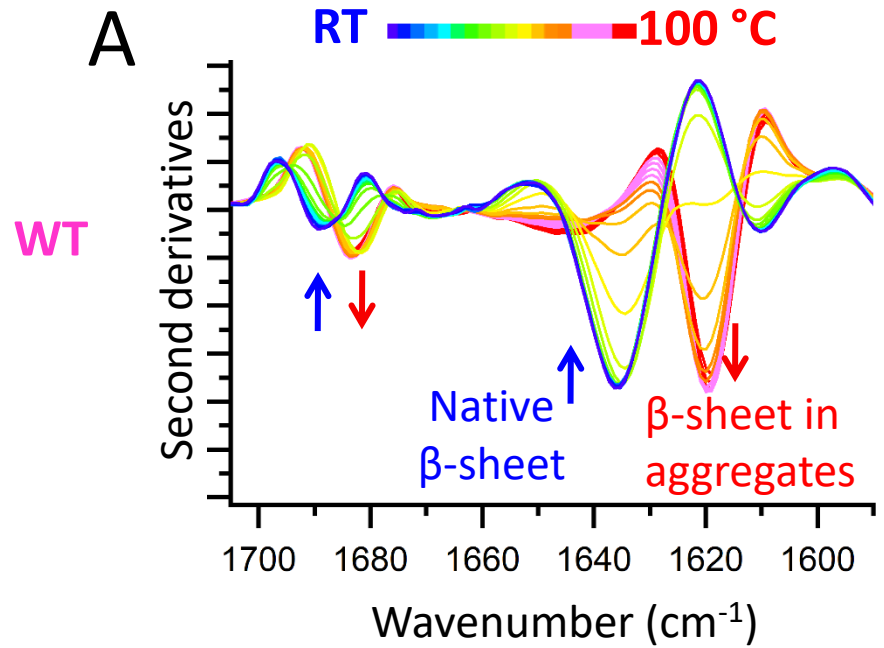


# Native $\beta$ 2m variants displayed identical secondary structures in solution as observed by FTIR spectroscopy



❖ The same secondary structures in solution

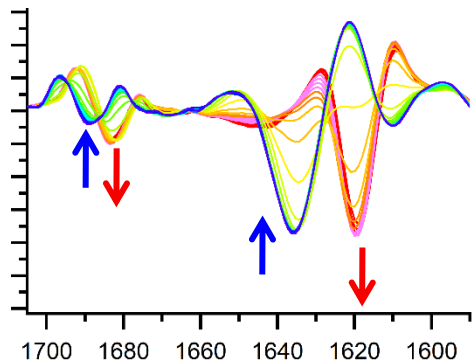
# Thermal stability and aggregation of $\beta 2m$ variants *in solution* by FTIR spectroscopy



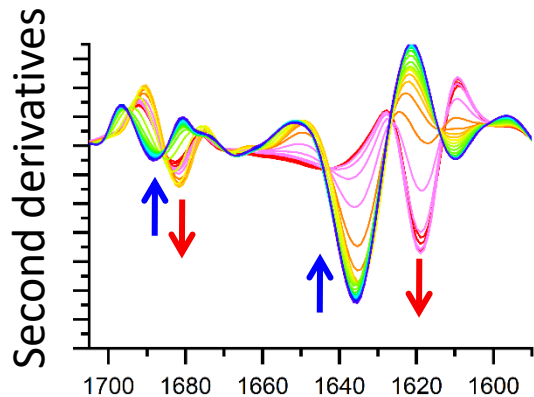
# Thermal stability and aggregation of $\beta 2m$ variants *in solution* by FTIR spectroscopy

RT  100 °C

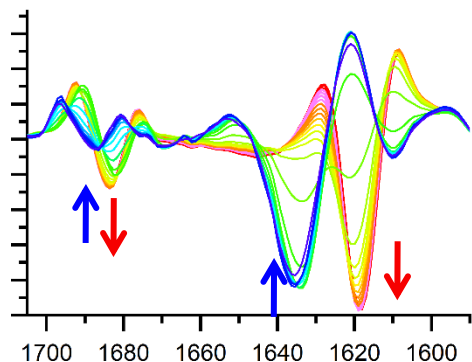
WT



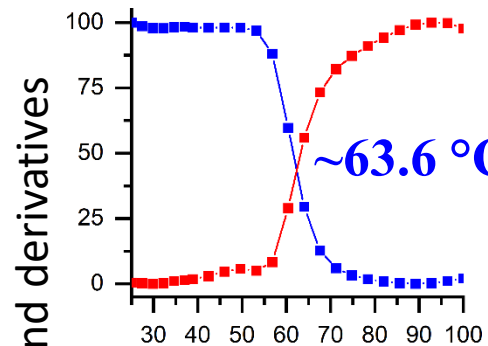
W60G




D76N

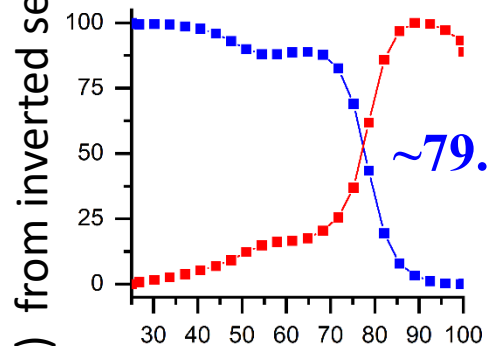


Wavenumber (cm<sup>-1</sup>)



 Native  $\beta$ -sheet  $\sim 1636 \text{ cm}^{-1}$

  $\beta$ -sheet in protein aggregates  $\sim 1615 \text{ cm}^{-1}$



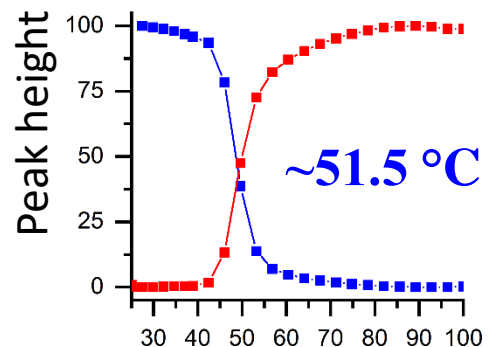
❖ The same secondary structures in solution

❖ Different thermal stability in solution



$T_{mp}$ :

W60G > WT > D76N

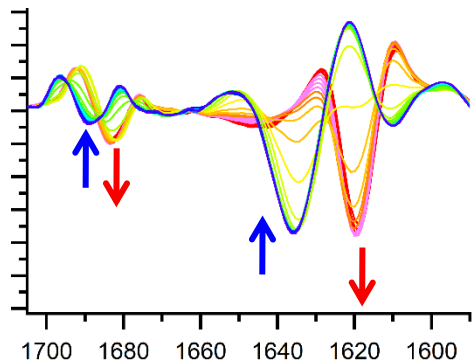


Temperature (°C)

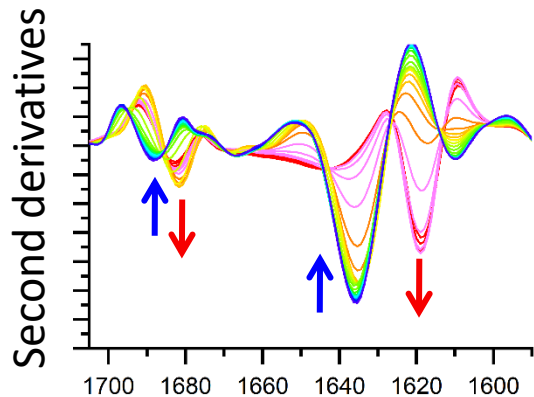
# Thermal stability and aggregation of $\beta 2m$ variants *in solution* by FTIR spectroscopy

RT  100 °C

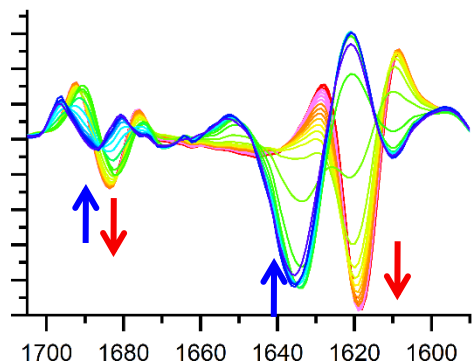
WT



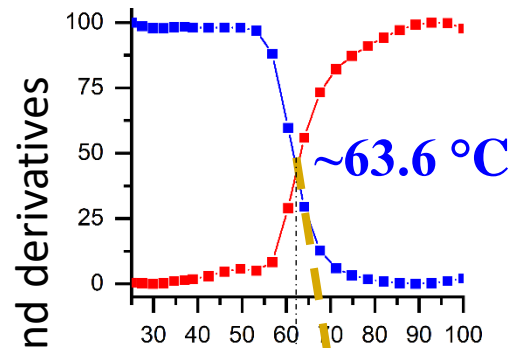
W60G



D76N



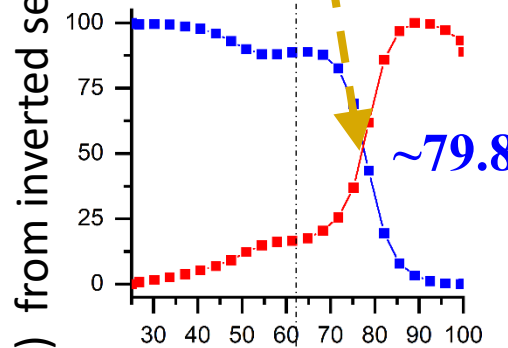
Wavenumber (cm<sup>-1</sup>)



 Native  $\beta$ -sheet  $\sim 1636 \text{ cm}^{-1}$

  $\beta$ -sheet in protein aggregates  $\sim 1615 \text{ cm}^{-1}$

$\sim 63.6 \text{ }^\circ\text{C}$



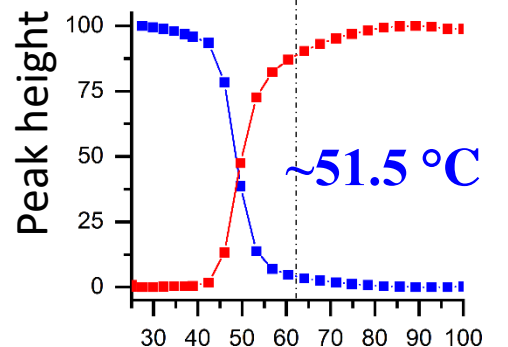
❖ The same secondary structures in solution



❖ Different thermal stability in solution

$T_{mp}$ :

W60G > WT > D76N



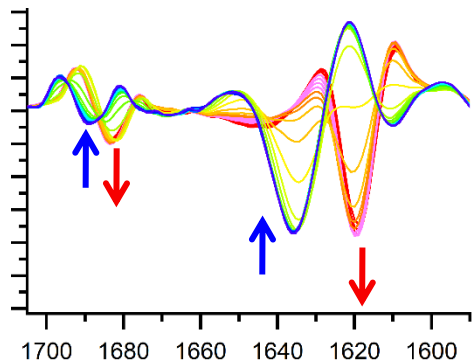
$\sim 51.5 \text{ }^\circ\text{C}$

Peak height (%) from inverted second derivatives  
Temperature (°C)

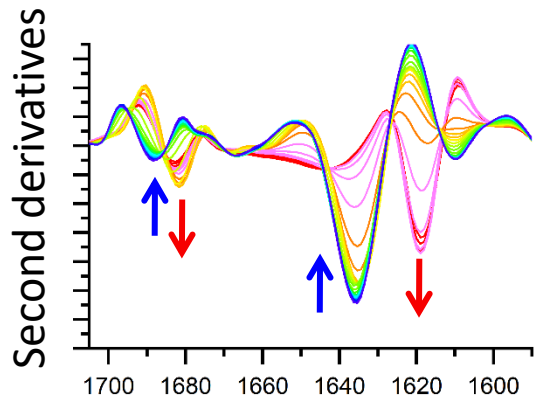
# Thermal stability and aggregation of $\beta 2m$ variants *in solution* by FTIR spectroscopy

RT  100 °C

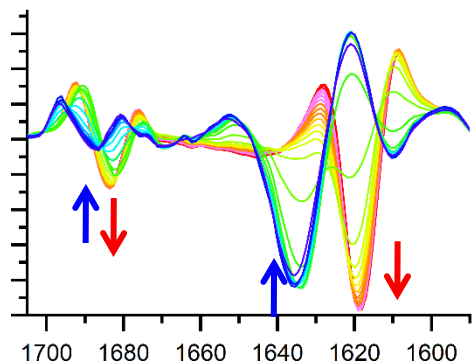
WT



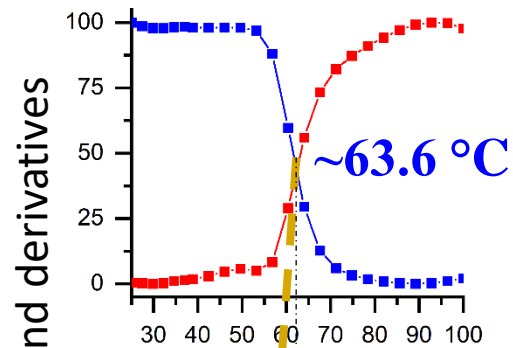
W60G




D76N

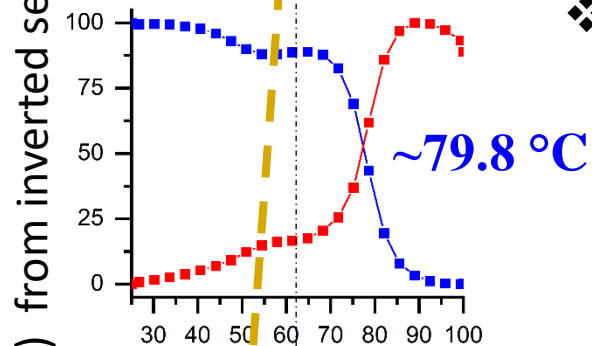


Wavenumber (cm<sup>-1</sup>)



 Native  $\beta$ -sheet  $\sim 1636 \text{ cm}^{-1}$

  $\beta$ -sheet in protein aggregates  $\sim 1615 \text{ cm}^{-1}$



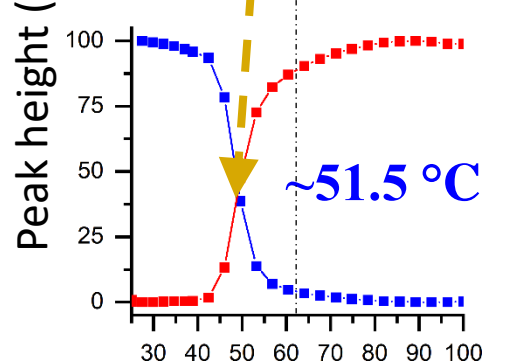
❖ The same secondary structures in solution

❖ Different thermal stability in solution



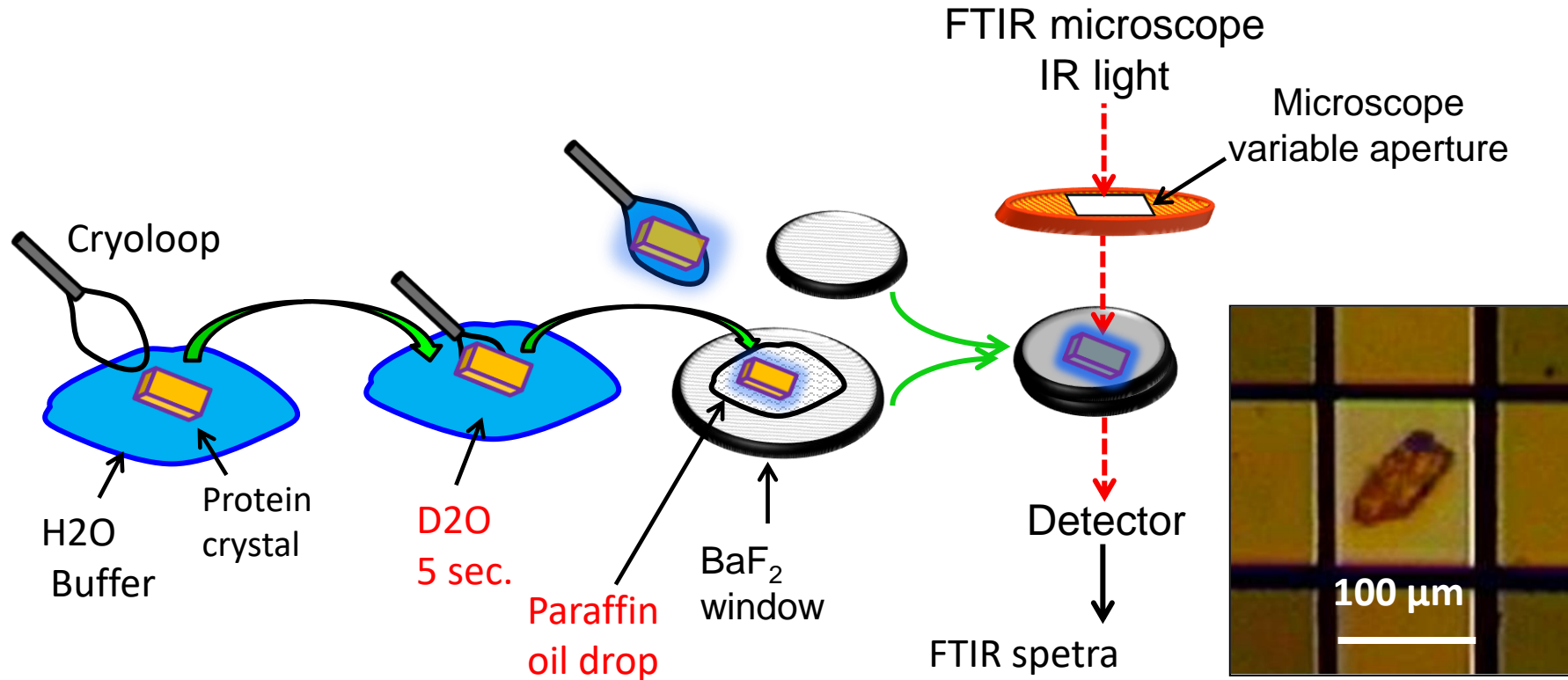
$T_{mp}$ :

W60G > WT > D76N



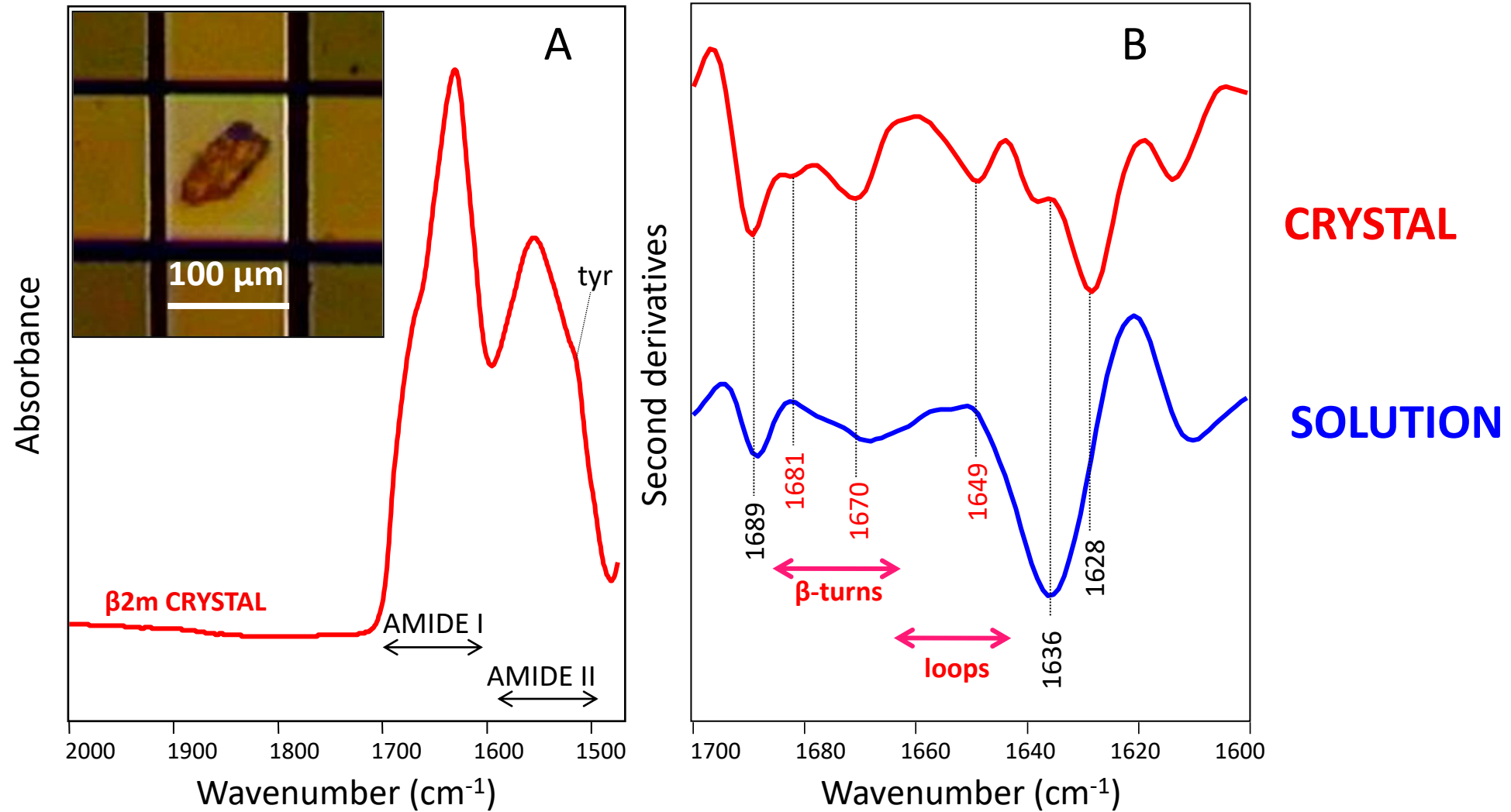
Temperature (°C)

# FTIR microspectroscopy of $\beta$ 2m single crystals





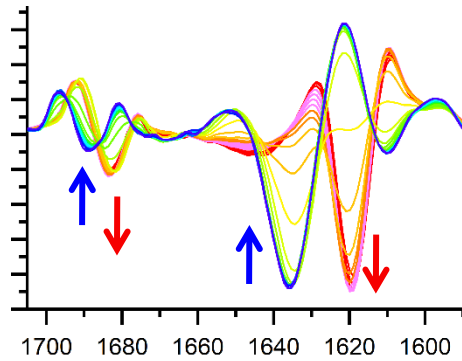
# FTIR microspectroscopy of wt $\beta$ 2m single crystals



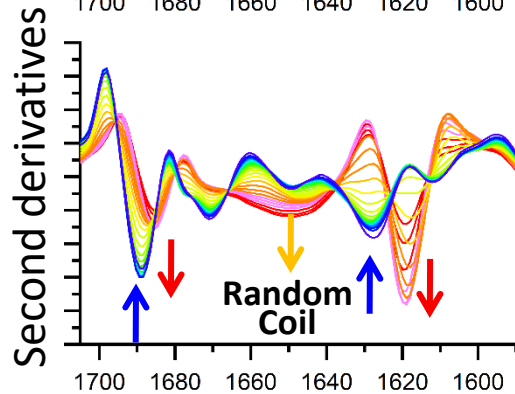
# Thermal stability and aggregation of WT $\beta$ 2m *in solution* and *in crystallo* by FTIR spectroscopy

RT  100 °C

Solution:  
0.4 °C/min

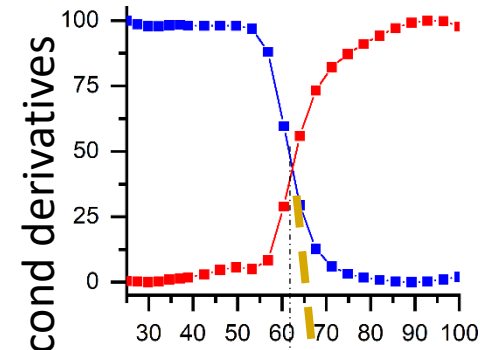
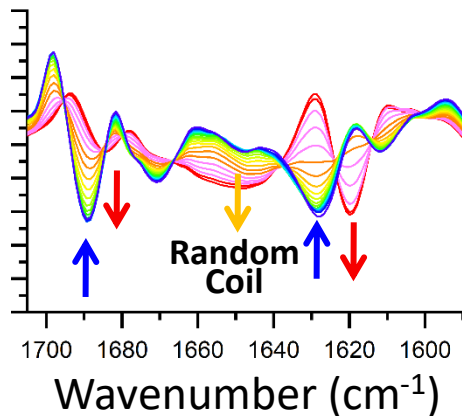


Crystal ensembles:  
0.4 °C/min



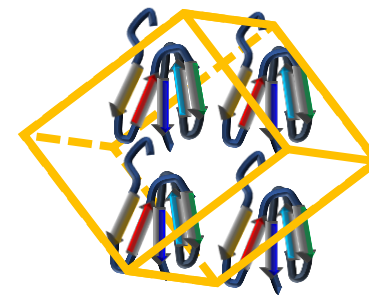
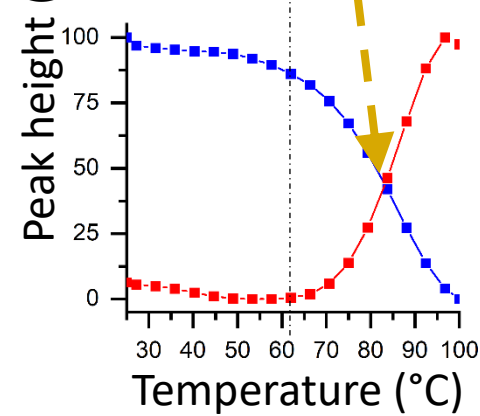
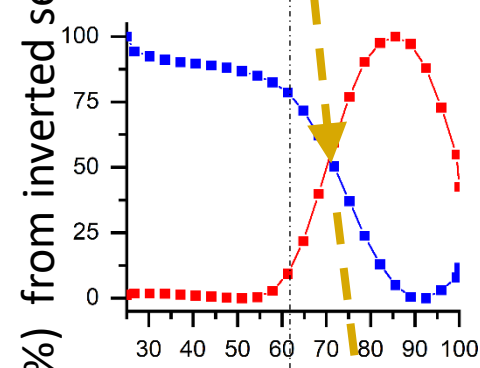
0.4 °C/min

1 °C/min

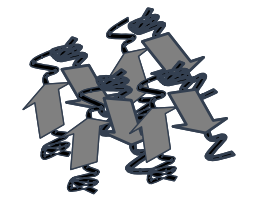


Native  $\beta$ -sheet  $\sim 1636 \text{ cm}^{-1}$

$\beta$ -sheet in protein aggregates  $\sim 1615 \text{ cm}^{-1}$



Crystal melting/protein unfolding and aggregation

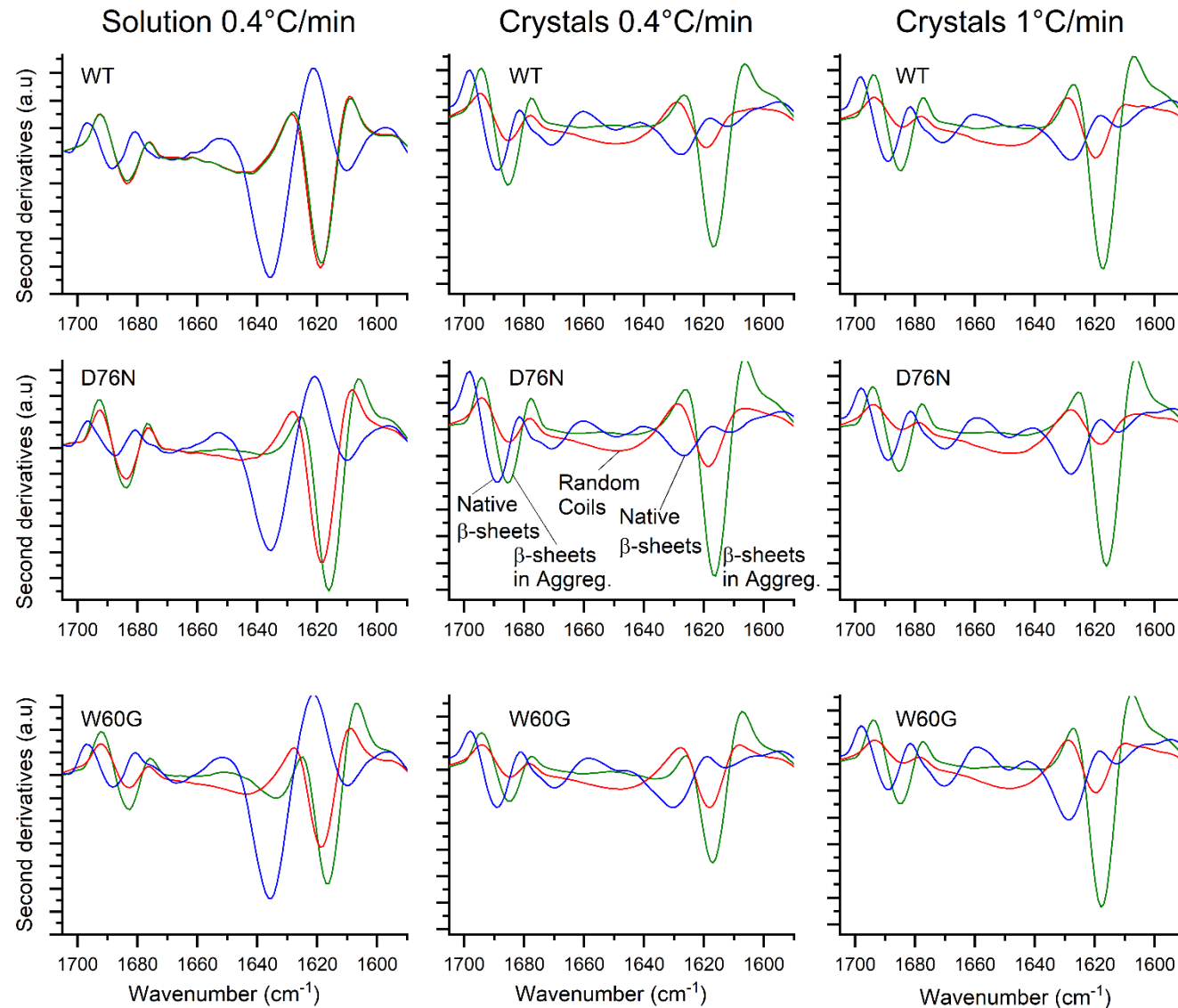


$\beta$ -sheet rich aggregates

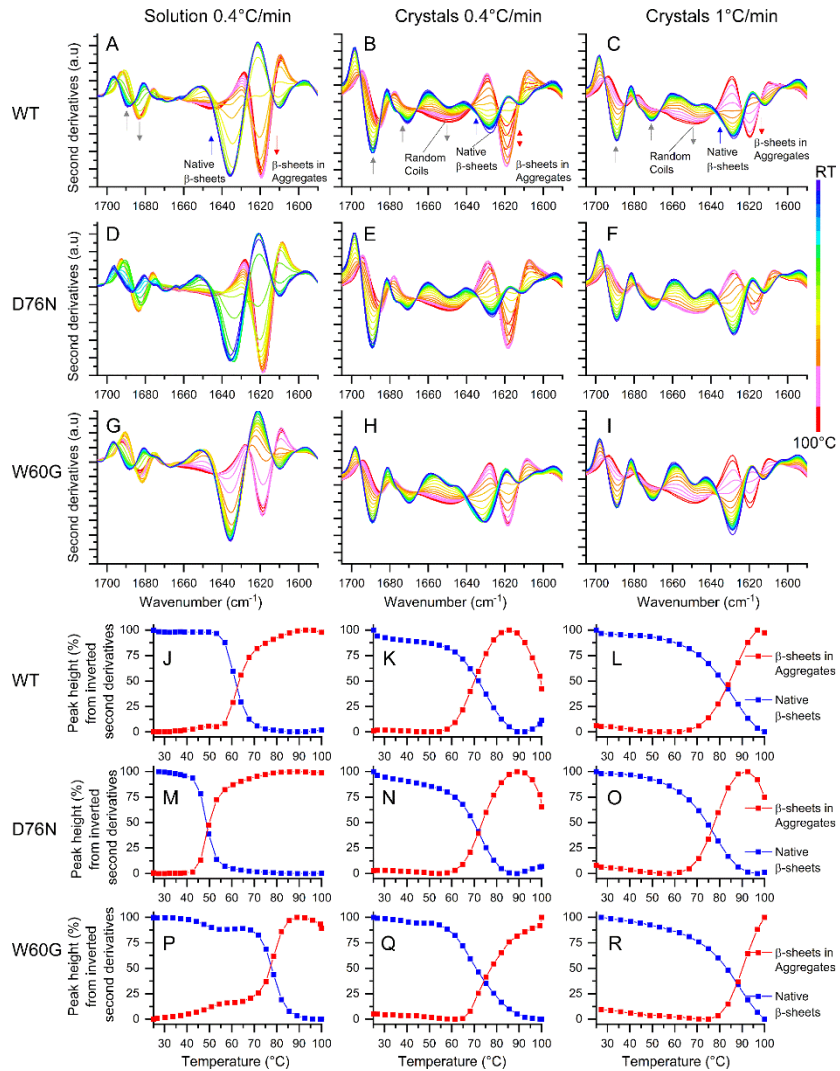
- ❖ Increased protein thermal stability *in crystallo* compared to *in solution*
- ❖ Heating rate affects  $T_{mp}$

# Aggregation of the $\beta 2m$ variants induced by thermal treatment

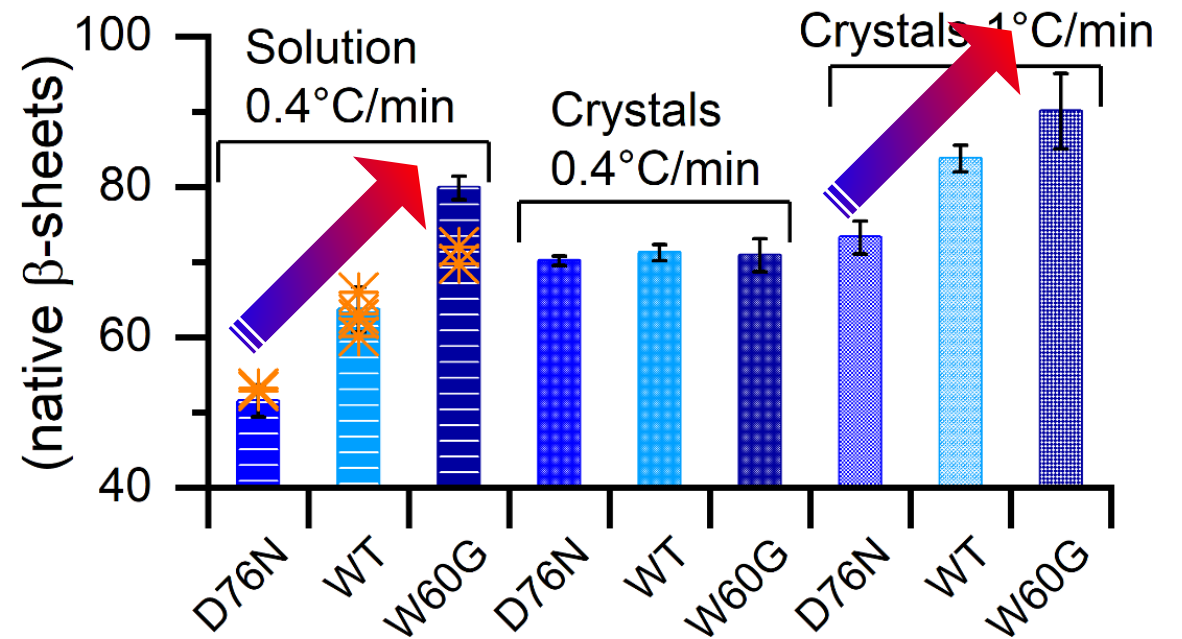
Room temperature  $\rightarrow$  100°C  $\rightarrow$  Room temperature



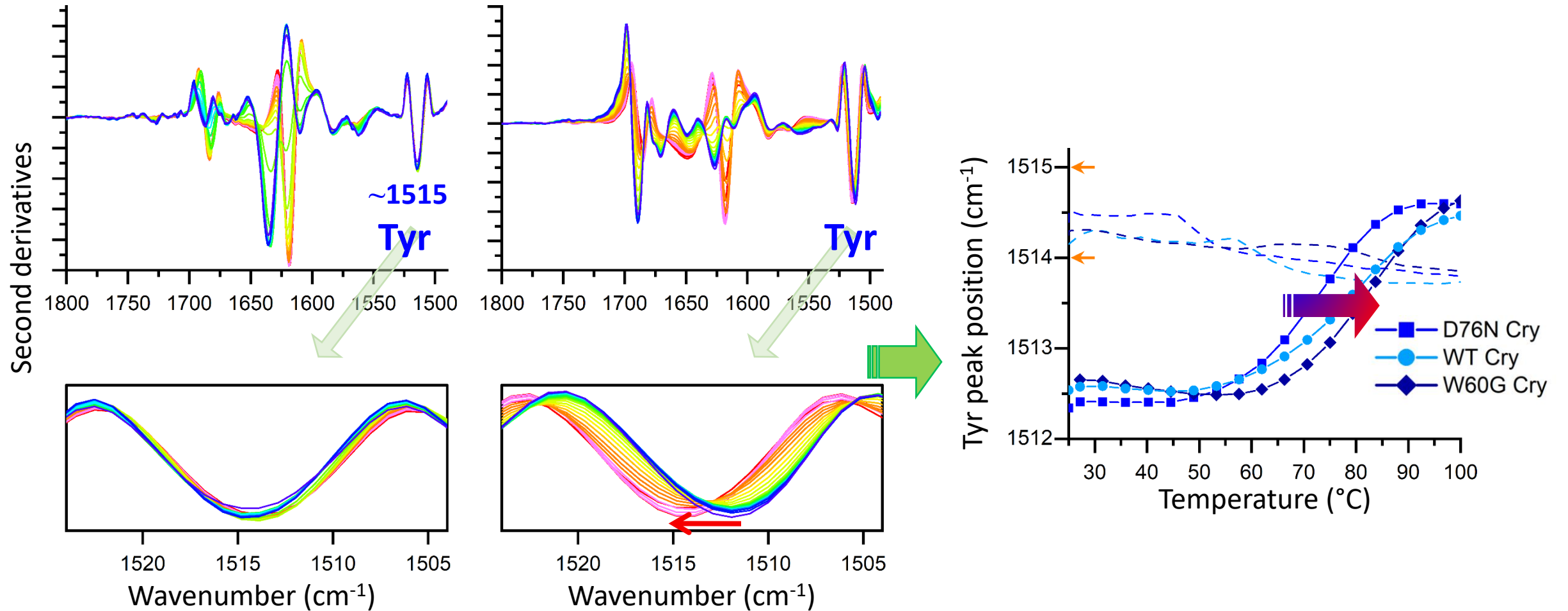
# Thermal stability of $\beta 2m$ variants assessed by FTIR spectroscopy



$T_{mp}$  ( $^{\circ}\text{C}$ )



# Temperature dependence of the tyrosine peak positions (ring $\nu(\text{CC})$ mode) of $\beta 2\text{m}$ variants in solution and in the crystalline states



# Conclusions

- Our data indicate that FTIR (micro-)spectroscopy allows
  - to study the secondary and tertiary structures of individual protein crystals and of crystal ensembles
  - to assess crystal stability by monitoring secondary and tertiary structure unfolding and protein aggregation simultaneously.
- FTIR spectra of crystals provide in general more information compared with the spectra measured in solution because of a higher number of well resolved Amide I components.
- The detailed comparison of protein stability using crystalline and solution samples indicates
  - a higher stability of the protein in the crystalline state compared to the solution state
  - the same stability trend for the protein in the crystalline state compared to the solution state.
- FTIR (micro-)spectroscopy, being a non-invasive approach, could be applied to the study of reactions, interactions, and protein conformational rearrangements within crystals, representing a promising tool to obtain structural information on protein crystals complementary to crystallographic analyses.



# Acknowledgements

University of Milano:

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A. Relini

Université de Lyon:

Le Marchand T  
Pintacuda G

University of Milano-Bicocca:

D. Ami



Thank you!